

RESEARCH ARTICLE

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Phylotranscriptomics points to multiple independent origins of multicellularity and cellular differentiation in the volvocine algae

Charles Ross Lindsey¹, Frank Rosenzweig^{1,2} and Matthew D. Herron^{1*} 

Abstract

Background: The volvocine algae, which include the single-celled species *Chlamydomonas reinhardtii* and the colonial species *Volvox carteri*, serve as a model in which to study the evolution of multicellularity and cellular differentiation. Studies reconstructing the history of this group have by and large relied on datasets of one to a few genes for phylogenetic inference and ancestral character state reconstruction. As a result, volvocine phylogenies lack concordance depending on the number and/or type of genes (i.e., chloroplast vs nuclear) chosen for phylogenetic inference. While multiple studies suggest that multicellularity evolved only once in the volvocine algae, that each of its three colonial families is monophyletic, and that there have been at least three independent origins of cellular differentiation in the group, other studies call into question one or more of these conclusions. An accurate assessment of the evolutionary history of the volvocine algae requires inference of a more robust phylogeny.

Results: We performed RNA sequencing (RNA-seq) on 55 strains representing 47 volvocine algal species and obtained similar data from curated databases on 13 additional strains. We then compiled a dataset consisting of transcripts for 40 single-copy, protein-coding, nuclear genes and subjected the predicted amino acid sequences of these genes to maximum likelihood, Bayesian inference, and coalescent-based analyses. These analyses show that multicellularity independently evolved at least twice in the volvocine algae and that the colonial family Goniaceae is not monophyletic. Our data further indicate that cellular differentiation arose independently at least four, and possibly as many as six times, within the volvocine algae.

Conclusions: Altogether, our results demonstrate that multicellularity and cellular differentiation are evolutionarily labile in the volvocine algae, affirming the importance of this group as a model system for the study of major transitions in the history of life.

Keywords: Phylotranscriptomics, Volvocine algae, Green algae, Phylogeny, Multicellularity

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Background

The evolution of multicellularity is widely considered a major transition in the history of life [1–4]. Multicellularity not only gave rise to most of the visible life forms on the planet, but also opened the door to cellular differentiation, including that between somatic and reproductive cells, a hallmark feature of sexual reproduction in eukaryotes that exhibit morphological complexity [3, 5, 6]. Questions regarding the evolution of multicellularity and cellular differentiation have been approached using the fossil record [7–9], laboratory evolution [10–13], and comparative approaches that include superimposing cell biology upon molecular phylogeny [14–16]. The last of these approaches is predicated on the assumption that the cell biology and molecular phylogeny are mutually informative, an assumption that requires the phylogeny itself to be accurate.

The volvocine green algae have proved especially useful for investigating the major transition leading to multicellularity. The group consists of ~ 50 extant species, which exhibit a range of body plans, cell numbers, sizes, and forms of sexual reproduction. The smallest of these are single-celled (e.g., *Chlamydomonas reinhardtii*); the largest, at up to 3 mm in diameter and up to 50,000 cells, are spheroidal, swimming colonies in the genus *Volvox*. Since the initial “very pleasant sight” of swimming *Volvox* colonies described by Van Leeuwenhoek more than 300 years ago [17], the volvocine algae have come to be accepted as a useful model system in which to address questions related to the origins of multicellularity and cellular differentiation [18, 19]. Multiple species have now had their genomes sequenced [20–24], and those of unicellular *C. reinhardtii* and multicellular *V. carteri* forma *nagariensis* are well-annotated [23, 24]. However, the volvocine algae encompass more than two organisms representing alternative forms of life in terms of size and development. Vegetative forms range in characteristic cell number from 1 to ~ 50,000 and exhibit intermediate degrees of complexity likely similar to extinct ancestors. Further, multicellularity and cellular differentiation arose within the volvocine algae much more recently than those traits arose in animals: ~ 220 million years ago [25] versus ~ 600 million years ago [26], respectively.

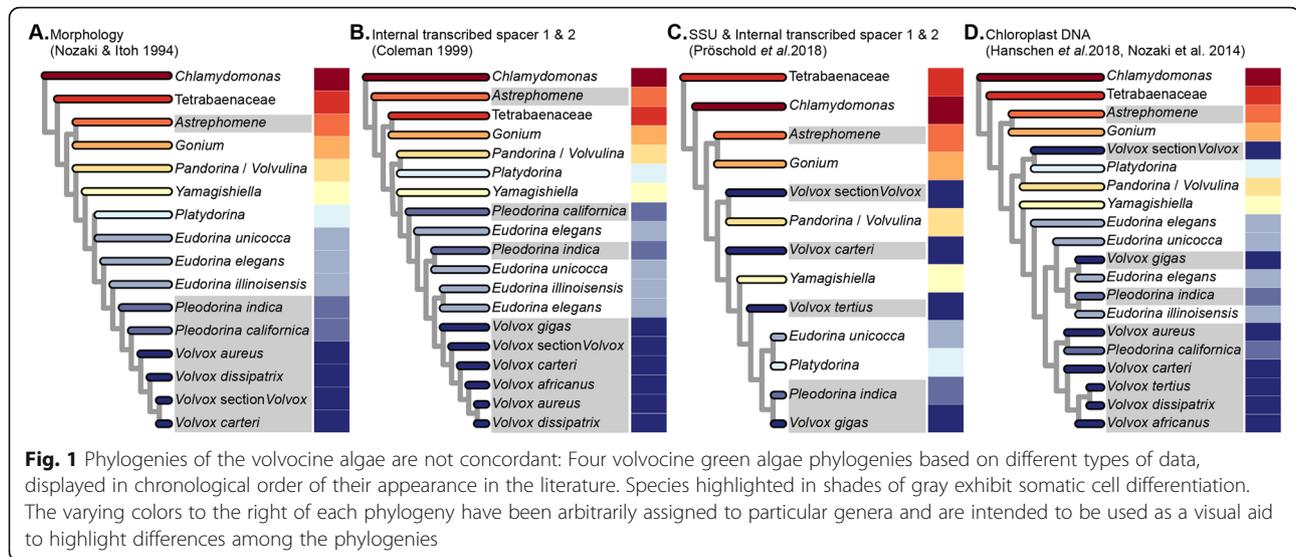
Evolution of the volvocine algae has sometimes been viewed as a linear progression in size and complexity [27, 28]. Unicellular taxa such as *Chlamydomonas* occupy one end of this continuum, while fully differentiated, multicellular taxa such as *Volvox* occupy the other. This concept, the “volvocine lineage hypothesis”, used a streamlined phylogeny of the volvocine algae to help explain how a multicellular species with complete germsoma differentiation such as *Volvox* might evolve from a unicellular, *Chlamydomonas*-like ancestor. However,

morphological and molecular phylogenetic studies suggest that the history of the volvocine algae may be more complicated, as cellular differentiation, different modes of sexual reproduction, and varying body plans appear to have evolved multiple times within the group [29, 30].

Current understanding of the major evolutionary relationships within this group has often been based on the analysis of five chloroplast gene sequences [14, 25, 31–35]. Chloroplast gene-based phylogenies have also been used to carry out ancestral-state reconstructions [14, 29, 30, 36], opening a window on how multicellularity and cellular differentiation evolved within the volvocine algae. Overall, the branching order of most chloroplast gene-based phylogenies is defined by two related groups: (i) a set of unicellular species (e.g., *Chlamydomonas reinhardtii*) that are paraphyletic with respect to (ii) a clade that encompasses the three major families of colonial volvocine algae: Tetrabaenaceae (*Tetrabaena* and *Basichlamys*), Goniaceae (*Gonium* and *Astrephomene*), and Volvocaceae (*Colemanosphaera*, *Eudorina*, *Pandorina*, *Platydorina*, *Pleodorina*, *Volvox*, *Volvulina*, and *Yamagishiella*) (Fig. 1a, d). In this scheme, the Tetrabaenaceae is a sister group to the clade formed by the Goniaceae and Volvocaceae. Although this framework only takes into account family-level relationships, several conclusions can be drawn. *First*, the colonial species form a clade. *Second*, each of the three families is monophyletic. *Third*, monophyly among the colonial species implies that multicellularity evolved only once within that group with no reversion to unicellularity.

Two recent studies have called into question the monophyly of the colonial volvocine algae (Fig. 1c). Pröschold et al. [37] based their inferences on two datasets: one consisting of SSU *rDNA* sequences plus internal transcribed spacer (ITS) sequences 1 and 2, the other consisting of ITS sequences alone. Nakada et al. [38] used a single-gene 18S *rRNA* dataset. Both studies inferred that the colonial species are paraphyletic with respect to certain unicells in the genera *Chlamydomonas* and *Vitreochlamys*.

The taxonomic status of the Goniaceae has also been called into question by studies (Fig. 1b) that indicate the group is either not monophyletic [39] or that there is low support for a sister relationship between *Astrephomene* and *Gonium* [33, 37, 38]. Moreover, a number of recent volvocine algal phylogenies leave uncertainty as to how many times cellular differentiation evolved within the group. Chloroplast sequence data suggest at least 3 independent origins of cellular differentiation: in *Astrephomene*, in *Volvox* section *Volvox* (sometimes referred to as *Euvolvox*), and in the *Eudorina*, *Volvox*, *Pleodorina* (EVP) clade (Fig. 1b–d). Within the EVP clade it is unclear whether cellular differentiation in



Pleodorina thompsonii, *Volvox gigas* and *V. powersii*, and *Pleodorina starrii* and *P. indica* arose independently from that in *V. carteri* (Fig. 1b–d).

The foregoing uncertainties highlight the need for a new and more robust molecular phylogeny of the volvocine algae. These uncertainties may arise from incomplete taxonomic sampling, limited genetic sampling, or both. While five volvocine algal species have had their genomes sequenced, most taxonomically comprehensive phylogenetic inferences about this evolutionarily important group have been constructed using relatively small datasets. Most consist of the sequence of five chloroplast genes [14, 31, 34, 40] representing an aggregate of ~6000 nucleotide positions. Others consist of small (≤ 6) multi-gene datasets consisting of chloroplast gene(s), ribosomal molecular markers, or both [37, 38]. Moreover, the use of chloroplast genes in phylogenetic reconstruction can be problematic because they are effectively a single linkage group, they vary little among recently diverged species [41], and they are at increased risk of incomplete lineage sorting due to the retention of ancestral polymorphisms [42, 43].

Of special concern is the observation that volvocine phylogenies inferred using chloroplast genes (Fig. 1d) conflict with those constructed using nuclear genes (Fig. 1b, c) [37–39]. While conflicts between chloroplast and nuclear phylogenies are not unusual [44–46], they do foster ambiguity.

Here, we seek to resolve volvocine relationships using taxonomically dense sampling of multiple, unlinked loci. We have adopted a phylotranscriptomic approach that uses a concatenated amino acid alignment of 40 nuclear protein-coding, single-copy genes. We sequenced whole transcriptomes of 55 strains encompassing 47 nominal species and used previously published RNA-Seq data for

9 strains and amino acid alignments for 4 strains that were shared with our group by the De Clerck laboratory. Our goal was to derive a robust phylogeny of the volvocine algae that would enable inferences about the evolution of multicellularity, cellular differentiation, sexual dimorphism, and other traits in this group. Our results represent the most taxonomically comprehensive phylogeny yet produced of the volvocine algae using a nuclear dataset, including all described genera and multiple representatives of all genera that are not monotypic. Our results show that the colonial species do not form a clade, that the Goniaceae are not monophyletic, and that multicellularity has independently evolved at least twice and cellular differentiation at least four times within the volvocine algae.

Results and discussion

De novo transcriptome data makes possible 40 single-gene alignments

We sampled 68 taxa representing all presumed major lineages of the colonial volvocine algae and 9 of their nearest unicellular relatives. Because the phylogenetic position of *Chlamydomonas reinhardtii* has recently been called into question [37, 38], we used a member of the Trebouxiophyceae, *Chlorella variabilis*, as an outgroup (Table 1). All described volvocine genera were included, with multiple species represented for every genus that is not monotypic. Truly comprehensive taxon sampling was not possible, since several described species, especially in the genus *Volvox*, are no longer available in culture collections. While our main focus was to resolve relationships within the colonial volvocine algae, our study included several closely related unicellular taxa from the genera *Chlamydomonas* and *Vitreochlamys* in

Table 1 List of taxa used in this study and summary of sequencing and assembly. Under Strain or Pubmed ID, “CC” refers to Chlamydomonas Culture Collection at the University of Minnesota (CC, USA), “F” refers to Culture Collection of Freshwater Algae at the Institute of Hydrobiology, Chinese Academy of Sciences (FACHB, China), “N” refers to National Institute for Environmental Studies (NIES, Japan), “S” refers to Culture Collection of Algae at the University of Göttingen (SAG, Germany), and “U” refers to The Culture Collection of Algae at the University of Texas at Austin (UTEX, USA). “QRPMK” and “TR” under RNA Extraction Method refer to QIAGEN RNeasy Plant Mini Kit and TRizol RNeasy, respectively. Strains assigned an asterisk represent data from previously published studies, with accession numbers shown in Supplementary Materials: Table S1 [21, 47]

Taxa	Strain or Pubmed ID	RNA extraction method	Total # of raw reads	Total # of clean PE reads	Total # of Contigs	N50	G/C %
<i>Astrephomene gubernaculifera</i>	N-418	QRPMK	61460551	44350080	78105	3894	0.61
<i>A. perforata</i>	N-564	QRPMK	63350620	46811342	72913	3613	0.61
<i>Basichlamys sacculifera</i>	N-566	QRPMK	66205749	48327288	134880	1945	0.69
<i>Colemanosphaera angeleri</i>	F 2363*		28119810	27000176	120763	3348	0.62
<i>C. charkowiensis</i>	F 2326*		27967726	26876865	103286	3382	0.62
	N-3383	TR	58777161	40550696	119848	2523	0.62
<i>Chlamydomonas debaryana</i>	S 11-55a	QRPMK	87455695	64552440	105096	2776	0.67
	S 70.81	QRPMK	50406749	42095108	103962	2858	0.64
<i>C. globosa</i>	S 81.72	TR	73820020	49419928	120062	2079	0.65
<i>C. moewusii</i>	S 11-16f	QRPMK	54728573	40436359	118833	2498	0.6
<i>C. reinhardtii</i>	CC-503*						
<i>C. schloesseri</i>	S 2486	QRPMK	70163393	49255021	64233	3094	0.66
<i>Chlorella variabilis</i> NC64a	20852019*						
<i>Eudorina cylindrica</i>	F 2322*		26442110	25262148	103026	2846	0.64
	N-722	QRPMK	44479173	36297801	105029	2927	0.64
<i>E. elegans</i>	F 2321*						
	N-456	TR	84549961	56756776	122975	2108	0.63
	N-458	QRPMK	67451343	46725012	131335	2944	0.65
	N-568	QRPMK	83433797	69539684	120487	3614	0.64
	N-717	QRPMK	47625429	37616922	89118	2094	0.64
	N-719	QRPMK	57157324	43565312	98180	2976	0.63
	N-720	QRPMK	38134498	31388413	95662	3699	0.61
<i>E. illinoisensis</i>	N-460	QRPMK	62485922	49396583	97014	2615	0.64
<i>E. minodii</i>	N-856	QRPMK	83574258	66413244	128802	4127	0.61
<i>E. peripheralis</i>	N-725	QRPMK	46607143	36528930	100934	2322	0.62
<i>E. unicocca</i>	S 24-1c	TR	54482446	37679863	110391	2267	0.62
<i>Gonium multicoccum</i>	N-737	QRPMK	65984190	44301578	122279	3275	0.64
<i>G. pectorale</i>	N-2863*						
<i>G. octonarum</i>	N-851	TR	60029472	39826211	104857	2302	0.65
<i>G. quadratum</i>	N-653	QRPMK	62431743	48230702	100468	3789	0.64
<i>G. viridistellatum</i>	N-654	QRPMK	43286980	31852410	97991	2837	0.65
<i>Pandorina colemaniae</i>	F 2361*		25181003	23823370	80843	3292	0.62
	N-572	TR	49862534	37589048	84053	2237	0.62
<i>P. morum</i>	F 2362*		29837807	28442854	147990	2674	0.61
	N-890	QRPMK	39337532	24490235	133742	3123	0.61
<i>Platydorina caudata</i>	N-728	QRPMK	48702213	36896556	125206	3503	0.61
<i>Pleodorina indica</i>	N-736	QRPMK	57564627	44082516	192570	2006	0.63

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Taxa	Strain or Pubmed ID	RNA extraction method	Total # of raw reads	Total # of clean PE reads	Total # of Contigs	N50	G/C %
<i>P. japonica</i>	U 2523	QRPMK	64897803	49495315	83662	3499	0.61
<i>P. starrii</i>	N-1362	TR	68790016	45935282	87641	2438	0.64
	N-1363	TR	59466876	44726861	128732	1695	0.63
<i>P. thompsonii</i>	N-4126	QRPMK	81077847	61986735	118940	4005	0.62
<i>Tetrabaena socialis</i>	N-571*		42302450	41232209	62014	1012	0.65
<i>Vitreochlamys aulata</i>	N-878	QRPMK	80710519	54651118	132115	2673	0.65
	S 80.81	QRPMK	41008694	32585611	84834	2100	0.65
<i>V. nekrassovii</i>	S 11-10	QRPMK	65658295	48341108	109360	3304	0.62
<i>V. ordinata</i>	N-882	TR	60631467	42981449	75434	2778	0.69
<i>Volvox africanus</i>	N-863	QRPMK	70739371	52378949	110066	4586	0.55
<i>V. aureus</i>	N-541	QRPMK	80821130	64315625	106245	4065	0.55
<i>V. barberi</i>	N-730	QRPMK	49805303	37197105	152910	2217	0.58
<i>V. carteri f. kawasakiensis</i>	N-732	TR	53994112	34239609	86004	1982	0.58
<i>V. carteri f. nagariensis</i>	20616280*						
	N-865	QRPMK	65624222	50641434	117314	3947	0.57
<i>V. carteri f. weismannia</i>	N-866	QRPMK	25913610	20987137	78230	2636	0.57
<i>V. dissipatrix</i>	N-4128	QRPMK	77137081	51045569	112190	5146	0.55
<i>V. ferrisii</i>	N-3986	QRPMK	49810357	37522413	91045	3281	0.57
<i>V. gigas</i>	N-867	QRPMK	50554912	37592146	151302	2499	0.63
<i>V. globator</i>	S 199.80	QRPMK	67413919	52861449	100861	4005	0.56
<i>V. kirkiorum</i>	N-543	QRPMK	44941633	34650729	89533	3081	0.56
<i>V. obversus</i>	N-868	QRPMK	67394604	50823546	99171	3881	0.58
<i>V. ovalis</i>	N-2569	QRPMK	52242003	35319700	107999	3643	0.53
<i>V. powersii</i>	N-4127	QRPMK	55232784	42410688	140237	2908	0.62
<i>V. tertius</i>	N-544	QRPMK	86222840	62563240	108551	5454	0.54
<i>Volulina boldii</i>	N-893	QRPMK	25665262	20161297	158265	1593	0.62
<i>V. compacta</i>	F 2337*		27543035	26374210	90261	3550	0.61
	N-582	QRPMK	53045384	38991781	112583	3380	0.63
<i>V. pringsheimii</i>	N-895	TR	82233959	55849890	128304	3122	0.62
<i>V. steinii</i>	S 90-1	QRPMK	46133638	37699635	89515	3635	0.62
<i>Yamagishiella unicocca</i>	F 2364*		30922957	29574420	64415	3609	0.63

order to provide better phylogenetic resolution of the volvocine algae as a whole.

The total number of raw reads generated from RNA sequencing for each species ranged from 25,665,262 to 87,455,695 reads with an average of 60,194,849 reads per species. After quality trimming of the raw reads (see “Methods”), the total number of clean paired-end reads ranged from 20,161,297 to 69,539,684 with an average of

44,416,935 reads per species (Table 1). From the RNA-seq data, we assembled a total of 40 single-gene alignments that were later concatenated to a single alignment representing an aggregate of 12,650 amino acids, equivalent to 37,950 nucleotide positions, with a total of 5972 parsimony-informative sites. Numbers of informative positions in the single-gene alignments ranged from 40 to 446. Trees inferred using maximum likelihood (ML),

Bayesian inference (BI), and coalescence-based (CB) analyses were generally well-supported with some topological differences between the ML and BI analyses relative to the CB analysis, as described below.

Our results conflict with prior volvocine algal phylogenies in four respects

First, we find that the colonial volvocine algae are paraphyletic with respect to some unicellular species. *Second*, monophyly of the family Goniaceae is not supported. *Third*, section *Volvox* is inferred to be sister to the remaining Volvocaceae. *Fourth*, cellular differentiation

independently arose at least four and perhaps as many as six times within the volvocine algae.

Colonial volvocine algae are not monophyletic

All three of our phylogenetic analyses indicate that the colonial volvocines are not monophyletic (Figs. 2 and 3); further, an approximately unbiased (AU) test strongly rejected monophyly for this group ($p = 2.82e-38$) (Additional file 1: Fig. S1a). These findings represent a major departure from earlier chloroplast gene-based volvocine phylogenies [14, 25, 31–34, 40, 48], phylogenies based on morphological

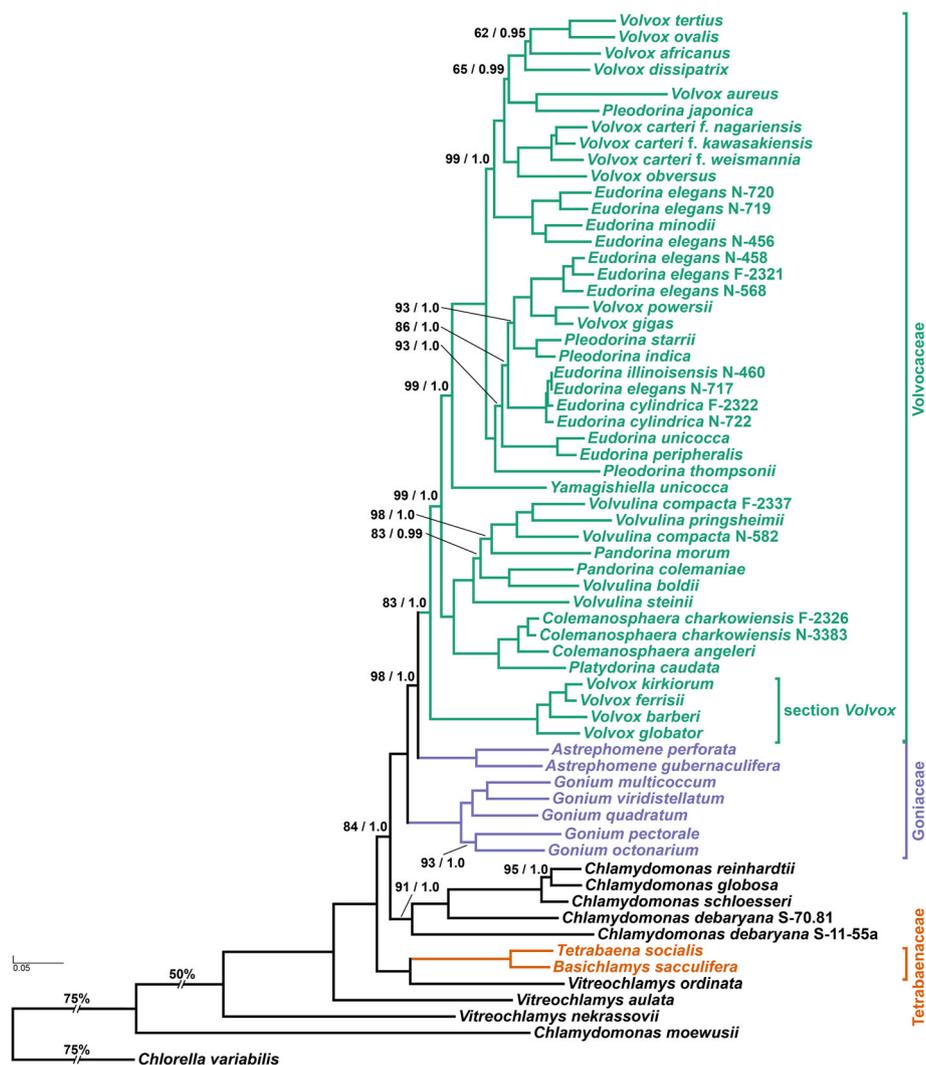
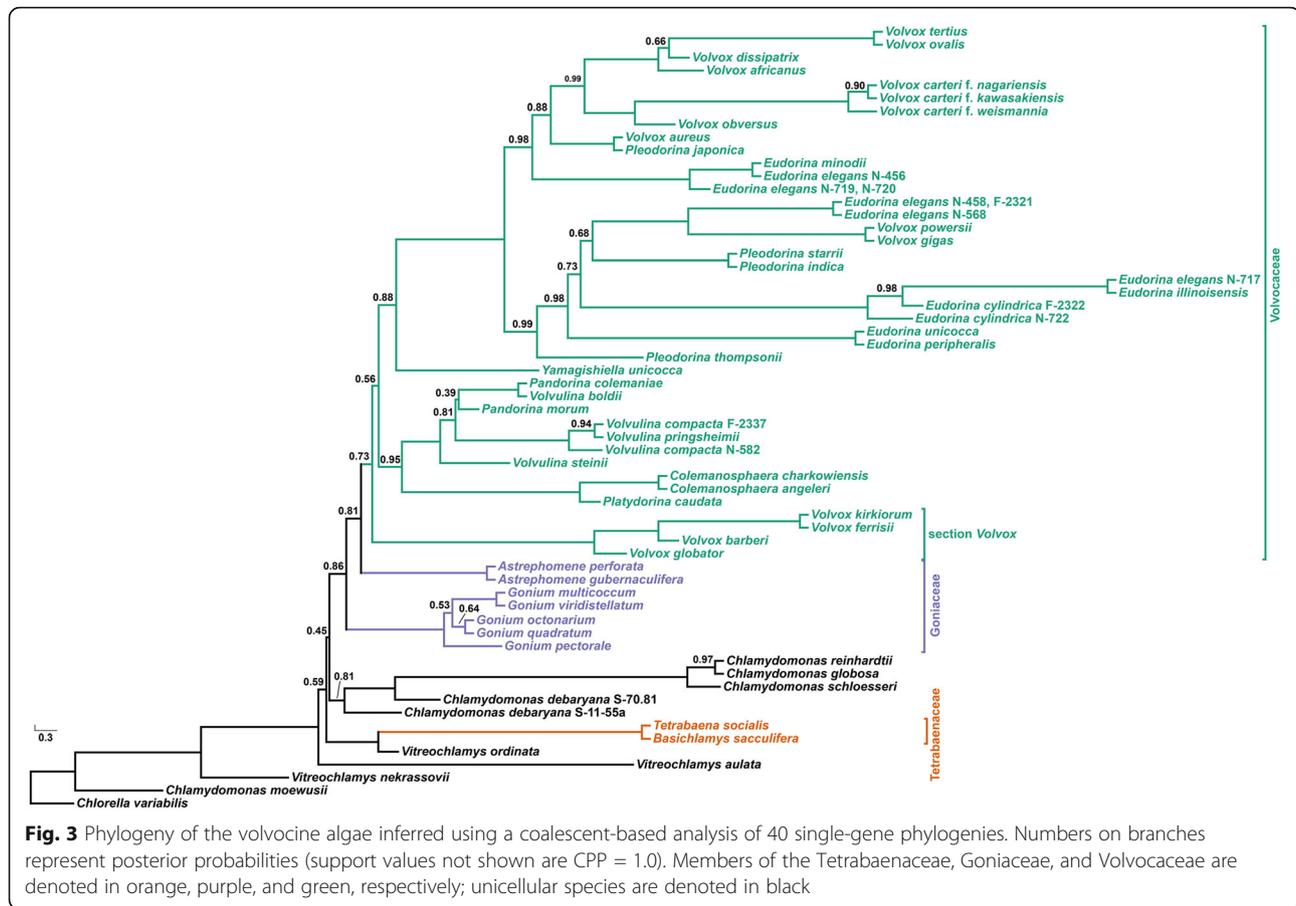


Fig. 2 Molecular phylogeny of the colonial volvocine algae (Tetrabaenaceae, Goniaceae, and Volvocaceae) and closely related unicellular taxa represented by *Chlamydomonas* and *Vitreochlamys* with *Chlorella variabilis* as the outgroup. The phylogenetic tree shown is based on a multi-gene dataset of single-copy, protein-coding nuclear genes (12,650 aligned amino acid positions of 68 taxa) inferred using the maximum likelihood method, the branching order of which is identical to that inferred in the Bayesian Inference using MrBayes. Numbers on branches represent bootstrap values and Bayesian posterior probabilities, respectively (all support values not shown are MLBS = 100, BPP = 1.0). Branch lengths correspond to genetic divergence, as indicated by the scale bar. Members of the Tetrabaenaceae, Goniaceae, and Volvocaceae are denoted in orange, purple, and green, respectively; unicellular species are denoted in black



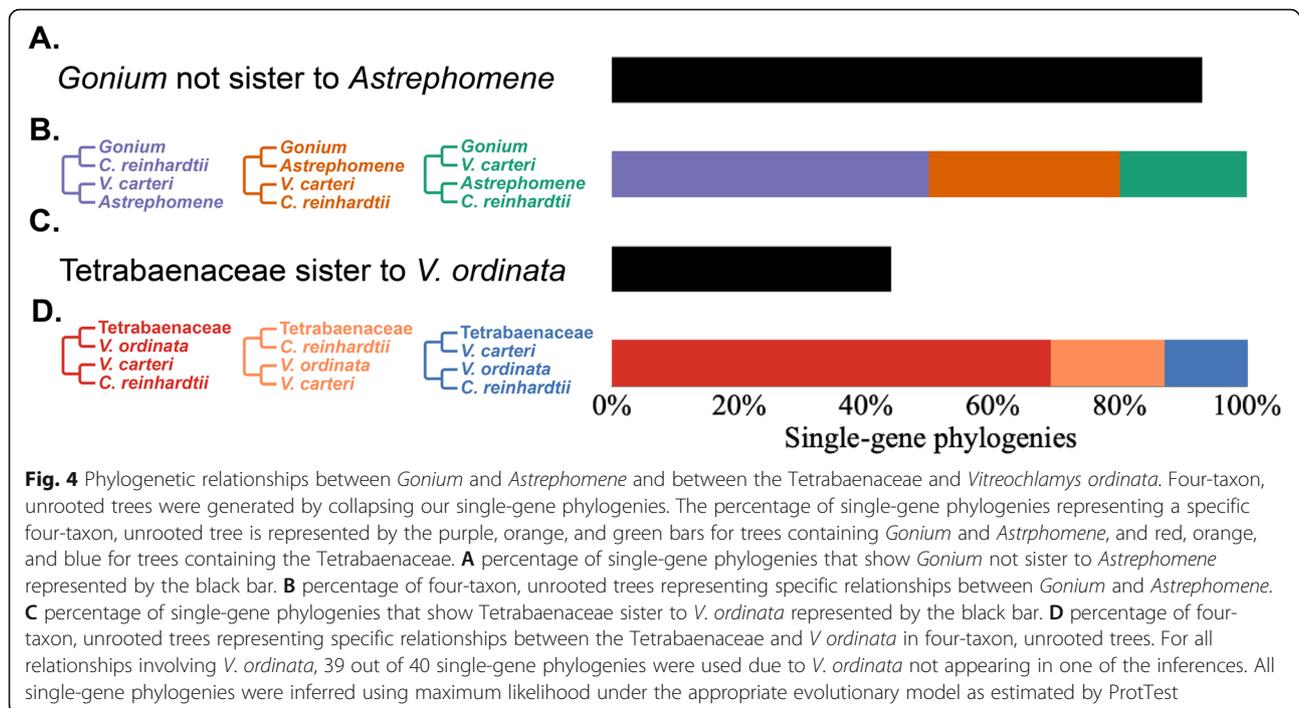
characters [49, 50], phylogenies inferred using ITS 1 and 2 sequences [39], as well as less taxonomically comprehensive phylogenies inferred using nuclear data [51], all of which suggest that the colonial volvocine algae are monophyletic.

Consistent with Pröschold et al. [37], our results support the view that multicellularity evolved independently in the Tetrabaenaceae and in the Goniaceae + Volvocaceae. In each analytical framework, the Tetrabaenaceae was found to be sister to *Vitreochlamys ordinata* rather than to the Goniaceae + Volvocaceae (Maximum Likelihood Bootstrap [MLBS] = 100, Bayesian Posterior Probabilities [BPP] = 1.0, Coalescent Posterior Probabilities [CPP] = 1.0). A sister relationship between the Tetrabaenaceae and *V. ordinata* was inferred in 17/39 of our single-gene phylogenies and in 27/39 of our 4-taxa, unrooted, single-gene phylogenies (Fig. 4). These results imply one independent origin of multicellularity in the Tetrabaenaceae and another origin in the Goniaceae + Volvocaceae.

Our results differ in key respects from a recent volvocine algal phylogeny inferred by Zhang et al. [51], which like ours is based on single-copy nuclear genes. Zhang et al. [51] sought to understand the evolutionary

relationships between two psychrophilic algae: *Chlamydomonas* sp. ICE-L and *Tetrabaena socialis* N-691. To do so, they constructed a phylogeny consisting of ICE-L, N-691, three colonial *Volvox* strains, and eight unicellular species, including *C. reinhardtii*. Among their conclusions was that *T. socialis* N-691 is sister to the Volvocaceae, which is at odds with results shown in Figs. 2 and 3. These results indicate that the Tetrabaenaceae is sister to *V. ordinata*, and together they are sister to *C. reinhardtii* + Goniaceae + Volvocaceae.

We hypothesized that the lack of concordance between our findings and those of Zhang et al. [51] could be attributed to limited taxon sampling. To test this hypothesis, we first confirmed that *T. socialis* N-691 and *T. socialis* N-571 are conspecific (Additional file 2: Confirming the conspecificity of *Tetrabaena socialis* N-571 and N-691) [52, 53]. Once we confirmed that N-691 and N-571 were conspecific, we were able to replicate the branching order produced by Zhang et al. [51] using our concatenated 40-gene dataset (Additional file 1: Fig. S2a) [51]. For our initial tree, we sampled our strains of *Chlamydomonas reinhardtii*, *C. moewusii*, *T. socialis*, *Volvox aureus*, *V. carteri* f. *nagariensis*, and *V. globator* to match taxa that were used in that study. For an



outgroup species, we sampled *Chlorella variabilis*. Multiple studies have shown that the accuracy of phylogenetic reconstruction can be improved by increasing the number of taxa sampled [54–56]. When we added more taxa and performed ML analysis on the new dataset, the three colonial volvocine families were no longer monophyletic. The Tetrabaenaceae were sister to *Vitreochlamys ordinata*, and this clade appeared sister to *C. reinhardtii* + Goniaceae + Volvocaceae (Additional file 1: Fig. S2b) [51]. These analyses confirm that the placement of *T. socialis* N-691 as sister to the Volvocaceae is an artifact of limited taxon sampling. From this, we draw three conclusions: *First*, the colonial volvocine algae are not monophyletic; *second*, at least two independent origins of multicellularity occurred within the volvocine algae; *third*, once multicellularity evolved no extant lineage reverted to the ancestral unicellular state (see Figs. 2 and 3).

The family Goniaceae is not monophyletic

Multiple volvocine phylogenies have concluded that the Goniaceae is monophyletic [14, 25, 29, 31, 33, 37, 38, 49, 50, 57, 58]. Our analyses suggest otherwise (Figs. 2 and 3): we find that *Astrephomene* is sister to the Volvocaceae (MLBS = 98, BPP = 1.0, CPP = 0.81) rather than to *Gonium*. This inference is strengthened by observations that 37/40 of our single-gene phylogenies show that *Gonium* and *Astrephomene* are not sister taxa, as do 20/40 of our four-taxon, unrooted phylogenies (Fig. 4). All three of our analyses indicate that *Astrephomene* is monophyletic and sister to the Volvocaceae clade (MLBS

= 98, BPP = 1.0, CPP = 0.81), with *Gonium* sister to *Astrephomene* + Volvocaceae (MLBS = 100, BPP = 1.0, CPP = 0.86). Furthermore, we performed an AU test where the monophyly of the Goniaceae was tested against our finding of parphyly for the Goniaceae. The null hypothesis, monophyly of the Goniaceae, was rejected ($p = 0.0446$) (Additional file 1: Fig. S1b). The inferred sister relationship between *Astrephomene* and the Volvocaceae is also consistent with the apparent synapomorphy of zygote germination producing a single gone cell, which is unique to these two taxa [50]

Prior studies have produced mixed results regarding monophyly of the Goniaceae, sometimes with low support values for the relevant relationships. Nozaki and colleagues [59] published four phylogenies inferred using a single chloroplast gene and different inference methods; all four trees either showed low support for monophyly of the Goniaceae or suggested a topology where *Astrephomene* is sister to *Gonium* + Volvocaceae. Coleman [39] inferred a volvocine phylogeny based on ITS-1 and ITS-2 sequences that showed *Astrephomene* sister to Tetrabaenaceae + *Gonium* + Volvocaceae; however, the bootstrap support for this suggested relationship was between 50 and 75%, indicating weak support for the branching order. Other phylogenies suggesting monophyly in the Goniaceae do so with weak or contradictory support [33, 37, 38].

Our inference that the Goniaceae are not monophyletic is consistent with some — but not all — of the analyses recently reported by Pröschold et al. [37] and Nakada et al. [38]. However, we should not disregard past

morphological and ultrastructural studies suggesting a close relationship between *Astrephomene* and *Gonium* [50, 60, 61]. These taxa differ from the Volvocaceae in that each cell, rather than the entire colony, is surrounded by a tripartite boundary [62]. This feature distinguishes their mode of colony formation from all other colonial algae within the Volvocaceae; our results suggest that it is ancestral to the Goniaceae + Volvocaceae and lost in the Volvocaceae.

Volvox section Volvox is sister to the remaining Volvocaceae

Our data indicate that *Volvox* section *Volvox* is not a subclade within either the *Pandorina* + *Volvulina* + *Colemanosphaera* (PVC) or *Eudorina* + *Volvox* + *Pleodorina*

(EVP) subclades. Older studies based on the *rbcL* chloroplast gene [49], ITS-1 and ITS-2 sequences [39], and morphology [50] suggest that section *Volvox* belongs to a clade that encompasses *Eudorina*, *Pleodorina*, and other *Volvox* species. More recent studies of the volvocine algae based on 5 chloroplast genes, or based on multiple datasets that include 1 chloroplast gene [37], suggest that section *Volvox* belongs to a clade that includes *Pandorina*, *Volvulina*, and *Platydorina* [14, 31], and (in the studies where it was included) *Colemanosphaera* [34, 40]. By contrast, all of our analyses indicate that section *Volvox* is monophyletic and sister to the remaining Volvocaceae (MLBS = 83, BPP = 1.0, CPP = 0.73). AU tests rejected the monophyly of section *Volvox* + *Colemanosphaera* + *Platydorina* (p-AU = 4.64e- 88) and the monophyly of section *Volvox* + the

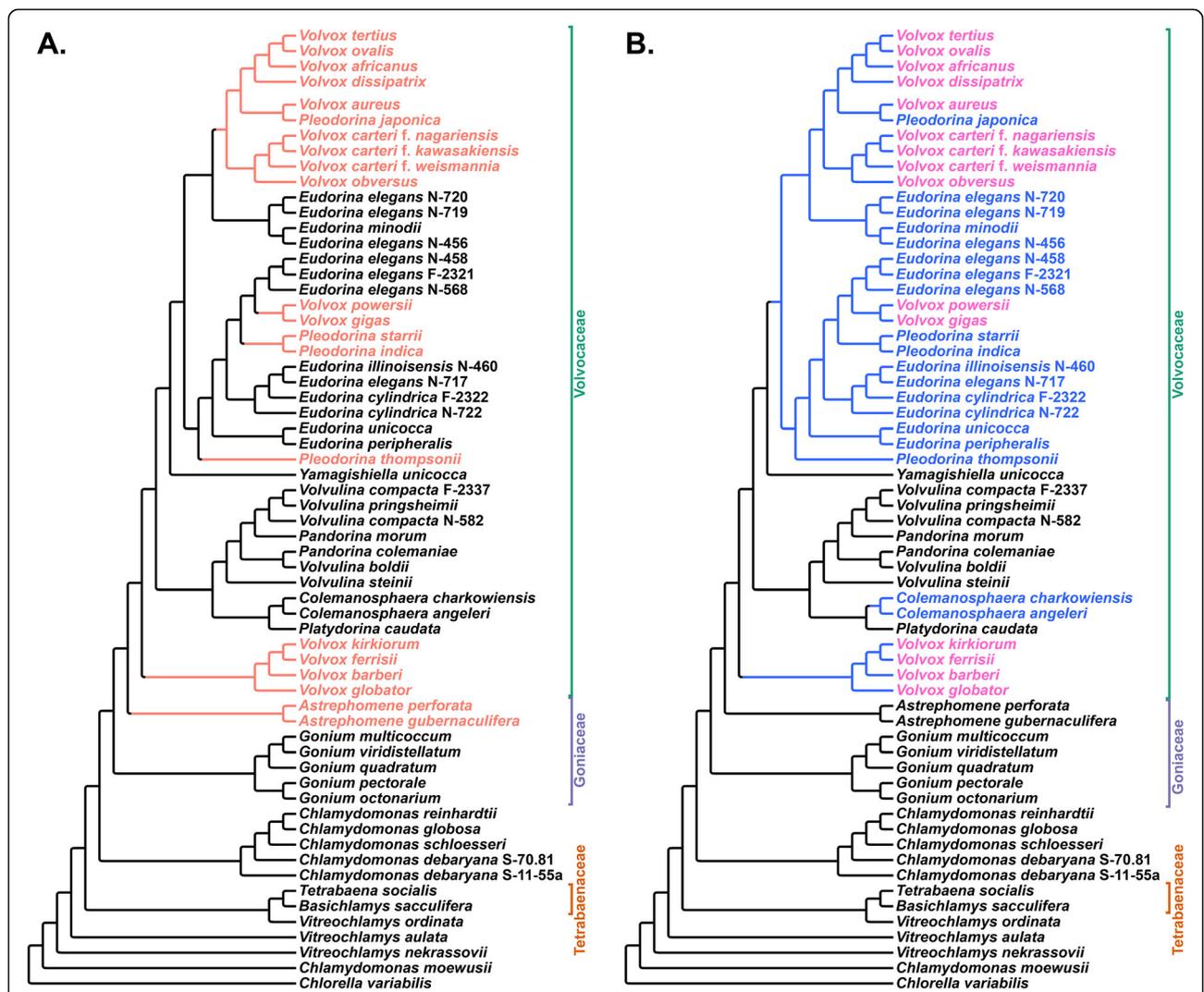


Fig. 5 **A** Phylogeny of the volvocine algae highlighting the lineages in which soma differentiation has evolved (peach). This tree indicates a minimum of four and maximum of six independent origins of cellular differentiation. **B** Phylogeny of the volvocine algae highlighting the lineages that are isogamous (black), anisogamous (blue), and oogamous (names in pink only). Both phylogenies were inferred using maximum likelihood

PVC clade (p-AU = 0.0332) (Additional file 1: Fig. S1c). These results bolster our finding that section *Volvox* is sister to the remaining Volvocaceae (Figs. 2 and 3).

Cellular differentiation independently arose at least four times in the volvocine algae

The last major difference between our results and earlier phylogenies concerns the number of independent origins of cellular differentiation. Prior literature suggests that cellular differentiation independently evolved at least three times: once in *Astrephomene*, once in section *Volvox*, and at least once in the EVP clade [14, 36]. By contrast, our results show a *minimum* of four independent origins of cellular differentiation: one in *Astrephomene*, one in section *Volvox*, and at least two in the EVP clade (Fig. 5a). We cannot exclude the possibility of two additional independent origins in the branches leading to *Pleodorina starrii* and *Volvox gigas* (Fig. 5a). In *Astrephomene*, section *Volvox*, *Pleodorina*, and *Volvox dissipatrix*, differentiated cells carry out the function of motility, whereas undifferentiated cells participate in both motility and reproduction [15]. The remaining *Volvox* species within the EVP clade have all evolved specialized germ cells for reproduction and somatic cells for motility [25, 30].

Isogamy is the ancestral mode of sexual reproduction

Consistent with past studies, our results suggest that isogamy, the production of similar sized, motile gametes, is the ancestral mode of sexual reproduction among the volvocine algae (Fig. 5b and Additional file 1: Table S2). Isogamy is present in the unicellular genera *Chlamydomonas* and *Vitreochlamys* and is retained within the multicellular genera *Astrephomene*, *Basichlamys*, *Gonium*, *Pandorina*, *Platydorina*, *Tetrabaena*, *Volvulina*, and *Yamagishiella*. *Colemanosphaera*, *Eudorina*, *Pleodorina*, and *Volvox* have all evolved either anisogamy or oogamy [34, 63–65]. Anisogamy appears to have independently evolved at least three times from an isogamous ancestor: in section *Volvox* and in both *Colemanosphaera* and EVP. Conventional anisogamy, which consists of two motile gamete types of unequal size, appears in *Colemanosphaera*, *Eudorina*, and *Pleodorina*. This finding differs from those of Hanschen et al. [29], who reported that anisogamy independently evolved twice among the volvocine algae from isogamous ancestors. Oogamy, a specialized form of anisogamy where the female gamete is immotile and significantly larger than the motile, male gamete, is inferred to have independently evolved at least three times in lineages leading to section *Volvox*, *V. gigas* + *V. powersii*, and in the clade containing *V. africanus*, *V. aureus*, *V. carteri*, *V. dissipatrix*, *V. obversus*, *V. ovalis*, and *V. tertius* [34, 63, 64]. This last finding confirms results from

Hanschen et al. [29], who also reported at least three independent origins of oogamy among the volvocine algae.

Platydorina caudata is sister to *Colemanosphaera*, and *Pandorina* is paraphyletic with respect to *Volvulina*

Within the PVC clade, our results add further support to the view that *Pandorina* is paraphyletic with respect to *Volvulina* (Figs. 2 and 3) [14, 25, 29, 33, 34, 39, 66]. Also, consistent with other multi-gene analyses *Colemanosphaera* appears to be monophyletic with high support (MLBS = 100, BPP = 1.0, CPP = 1.0) and sister to *Platydorina* (MLBS = 100, BPP = 1.0, CPP = 1.0) (Figs. 2 and 3) [31, 34, 35].

The genera *Eudorina*, *Volvox*, and *Pleodorina* are polyphyletic

Yamagishiella unicocca is sister to the *Eudorina*+*Volvox*+*Pleodorina* (EVP) clade, which encompasses two large subclades (MLBS=99, BPP=1.0, CPP=0.88) (Figs. 2 and 3). Our results support prior work suggesting that the genera *Volvox*, *Eudorina* and *Pleodorina* are not monophyletic [14, 25, 29–31, 33–36, 39, 67, 68]. The genus *Volvox* appears to be polyphyletic, with members represented across the two EVP subclades and the section *Volvox* clade. Members of both the *Pleodorina* and *Eudorina* genera are inferred to be polyphyletic across the two EVP subclades.

Historically, the genus *Volvox* has been divided into 4 sections – *Copelandosphaera*, *Janetosphaera*, *Merrillosphaera*, and *Volvox* – based on morphological [69] and molecular data [67]. A recent section-level revision of the genus *Volvox* [35] resulted in the creation and deletion of sections *Besseyosphaera* and *Copelandosphaera*, respectively. Hereafter, we will only refer to the revised taxonomic sections proposed by Nozaki et al. [35], with which our maximum likelihood, Bayesian inference, and coalescent-based results are in agreement (Additional file 1: Fig. S3) [35]. Our coalescent-based analysis suggests that each of the four sections is monophyletic, and that none encompass novel taxa not listed by Nozaki et al. [35] (Fig. 3). The branching order of our ML and BI analyses, however, suggests that section *Merrillosphaera* is not monophyletic (Additional file 1: Fig. S3) [35]. Our ML and BI analyses indicate that *V. africanus*, *V. dissipatrix*, *V. ovalis*, and *V. tertius* form a clade with *V. aureus* and *P. japonica* that is separate from the other *Merrillosphaera* taxa (MLBS=65, BPP=0.99) (Additional file 1: Fig. S3) [35]. In contrast, our CB analysis provides strong support (CPP=0.99) for the inference that the *Merrillosphaera* species are monophyletic (Fig. 3). Heeding our support values rather than only the branching order, we propose that the taxonomic system of the genus *Volvox* as outlined by Nozaki and colleagues [35] be retained.

Unicellular taxa are nested within the clade containing the colonial volvocine algae

Of the unicellular taxa, *Chlamydomonas debaryana*, *C. globosa*, *C. reinhardtii*, *C. schloesseri*, and *Vitreochlamys ordinata* are nested within the clade containing the colonial volvocine algae. Our results confirm prior studies showing the genus *Vitreochlamys* to be polyphyletic [38, 48]. The closest unicellular relative to the clade that contains the colonial algae + *C. reinhardtii* is suggested to be *V. aulata* (Figs. 2 and 3). This suggests that at least some members of *Vitreochlamys* are very closely related to the colonial volvocine algae. This relationship had been previously suggested by other studies [38, 70] including Nakazawa et al. [48], whose ultrastructural studies uncovered striking similarities in how these taxa formed pyrenoids and eyespot apparatus (stigma), and established their tripartite cell walls.

Chlamydomonas is a polyphyletic genus [20, 38, 71, 72] composed of at least 500 species [72]. Although we sampled only a handful of *Chlamydomonas* species, our data support this view and broadly agree with the *Chlamydomonas* relationships inferred by Pröschold et al. [37], who used a combination of molecular phylogenetic analyses, sporangium wall lysis tests, and ultrastructural analyses. Our data strongly support *C. schloesseri* being sister to *C. reinhardtii* + *C. globosa* (MLBS = 100, BPP = 1.0, CPP = 1.0) and designating *C. schloesseri* as a “true” *Chlamydomonas* species, as suggested by Pröschold et al. [37]. Our study is also in agreement with a recent study by Craig et al. [20] that shows *C. schloesseri* being sister to *C. reinhardtii* + *C. globosa*. Also, like Pröschold et al. [37], our analyses indicate that *C. debaryana* SAG 70.81 is sister to *Chlamydomonas schloesseri* and its relatives (MLBS = 100, BPP = 1.0, CPP = 1.0). However, unlike the Pröschold et al. [37] study, which proposed that strain *C. debaryana/Edaphochlamys debaryana* (SAG 11-55a) is sister to the Tetrabaenaceae, our analyses support the view that *C. debaryana/Edaphochlamys debaryana* is more closely related to *C. reinhardtii* (MLBS = 91, BPP = 1.0, CPP = 0.81) than to the colonial algae. Our finding is further supported by Craig et al. [20] who inferred that *C. debaryana/Edaphochlamys debaryana* + *Chlamydomonas sphaeroides* is sister to the clade containing *C. schloesseri* + *C. reinhardtii* + *C. globosa*. Our placement of *C. debaryana* (SAG 11-55a) could be a result of limited (N = 6) sampling within the *Chlamydomonas* genus, which was more extensively sampled by Pröschold et al. [37] (N > 30). Consistent with a prior study, *C. moewusii* appears to be more distantly related to the colonial volvocines than is *Vitreochlamys nekrassovii* [14].

Conclusions

Using a 40-protein dataset, we have shown that the Tetrabaenaceae and the Goniaceae + Volvocaceae likely represent two independent origins of multicellularity and that cellular differentiation has independently evolved at least four, and possibly six times within the volvocine algae. The separate origin of multicellularity within the Tetrabaenaceae highlights the need for certain volvocine genomes, such as *Vitreochlamys ordinata*, to be sequenced, assembled and annotated. Because *Vitreochlamys ordinata* is the unicellular sister taxon to the multicellular Tetrabaenaceae, detailed analysis of its genome could give future researchers insight into how the simple form of multicellularity observed among the Tetrabaenaceae might have evolved.

Our results suggest that both multicellularity and cellular differentiation are evolutionarily labile traits within the volvocine algae. We have established a robust phylogeny of this group, which we hope will assist future efforts aimed at re-evaluating ancestral character states and understanding the origins of multicellularity and cellular differentiation in the volvocine green algae. The fruit of such efforts could then be used to carry out ancestral-state reconstruction of traits related to cellularity, differentiation, and gamete size as well as to discern the evolutionary history of gene families across the volvocine algae as a whole and within its major clades.

Methods

Strains and culture conditions

Algal strains used in this study were obtained from the National Institute for Environmental Studies (NIES, Japan), the Culture Collection of Algae at the University of Göttingen (SAG, Germany), and the Culture Collection of Algae at the University of Texas at Austin (UTEX, USA). Strain provenance and culture collection ID numbers are shown in Table 1, with previously published data designated with an asterisk. All cultures were grown at 20–26 °C under cool-white LED lamps (4300K) with an intensity of 2500–2700 lux under a 14-h light/10-h dark cycle. A detailed description of each strain’s morphology, degree of cellular differentiation, and gamete size, as well as the medium used to culture each strain is provided in Additional file 1: Tables S2 and S3 [73–77], respectively.

RNA extraction procedures

Two protocols were used to isolate total RNA: a modified version of the TRizol RNeasy method described by Matt and Umen [78] and a slightly modified QIAGEN RNeasy Plant Mini Kit protocol. For a detailed description of each, please see Additional file 2: RNA extraction

procedures. Information on the protocol used for each strain is provided in Table 1.

Library preparation and sequencing

Before generating a sequencing library, RNA quality and quantity were assessed by Nanodrop and Qubit (Thermo Fisher Scientific, Waltham, MA 02451 USA). RNA integrity was evaluated using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA 95051, USA). mRNA was isolated using poly T beads, whereafter Illumina libraries were prepared using the NEBNext Ultra II Directional RNA Library Prep Kit. Library concentrations were determined fluorometrically; sequencing was carried out on the Illumina NovaSeq 6000 platform (Illumina, Inc., San Diego, CA 92122 USA) to generate 151 bp paired-end reads.

Quality control of reads

Raw read quality was assessed through FastQC v.0.11.8 with an additional FastQC assessment post-trimming. Quality control of the raw reads was completed with Trimmomatic v.0.39 [79] where the bases at the 5' and 3' end of each read are trimmed if found to be below a quality score of 3. A 4-base sliding window approach was used to trim the rest of the read once average quality fell below a score of 15; reads that were below a minimum length of 36 bases were discarded (LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36). If adapter content was detected by FastQC the additional ILLUMINACLIP step was used with the "TruSeq3-PE-2.fa" file provided by the Trimmomatic developers. If performed, the following ILLUMINACLIP parameters were used: 2:30:10 at the beginning of each command line. This allows for 2 "seed" mismatches where the seed is a short segment of the adapter that is being aligned in every section of the read. If more than 2 mismatches occurred, no trimming of the read occurred. Additionally, there had to be at least 30 matched bases in the paired-end palindrome read alignment and at least 10 matched bases between an adapter sequence and read.

De novo assembly

SOAPdenovo-Trans v1.0.4 [80] was used to assemble de novo transcriptomes from the quality filtered, paired-end reads using a k-mer size of 25 (SOAPdenovo-Trans-31mer all -s <config input file> -o <outfile> -K 25). Gap-Closer from the SOAPdenovo package was utilized to close gaps in each transcriptome using the same configuration file, which contains read-specific information and file paths, from the previous step (-b <config file> -a <.scafSeq file output by SOAPdenovo-Trans> -o <outfile> -l <max read length, int value> -t <thread number>). Default parameters were used for CD-HIT v4.8.1

[81] to reduce redundant transcripts from our de novo transcriptomes.

Orthologous gene identification for phylotranscriptomic analysis

The evolutionary history of the volvocine algae dates back at least 200 million years [25]. Over this timescale nucleotide sequences become saturated with substitutions, diminishing their phylogenetic utility [82]. Amino acid sequences were therefore chosen for our alignments, as they are known to be more reliable for ascertaining distant evolutionary relationships [83]. De Clerck and colleagues identified 58 nuclear protein-coding, single-copy genes that were members of highly conserved gene families across the green algae (*Chlorophyceae*, *Prasinophytes*, and *Trebouxiophyceae*) and land plants (*Streptophyta*) [84]. Their amino acid alignment of the 58 nuclear protein-coding genes that includes *Chlamydomonas reinhardtii* CC-503, *Chlorella variabilis* NC64A, *Gonium pectorale* NIES-2863, and *Volvox carteri* HK10 was kindly shared with our research team. Out of the 58 genes shared, we used 40 for our gene alignments. In order to identify those specific genes in the de novo transcriptomes of our taxa, a Basic Local Alignment Search Tool (BLAST) server was established in our lab, and a unique BLAST database for each taxon was created following the instructions in the BLAST manual. A BLASTP search using the *C. reinhardtii* CC-503, *G. pectorale* NIES-2863, and *V. carteri* HK10 genes from De Clerck et al. [84] as our query sequences enabled us to identify the orthologous genes for each of our taxa.

Gene sequence alignments and phylotranscriptomic analysis

The BLASTP results were used to identify the scaffold and open read frame where each gene was located in a strain's transcriptome. Using a custom Python script (Additional file 3), each scaffold was extracted from its transcriptome and translated in the appropriate reading frame; then, the translated scaffold was added to an alignment file. For consistency, we generated de novo transcriptomes since we lacked a reference genome for most of our sequenced strains. At times, a gene was found to be incomplete for a given taxon due to assembler or sequencing error after manual examination. When this was determined to be the case, the gene was manually stitched together. This was done in a highly conservative manner: if we could not ascertain whether or not a gene was incomplete due to assembler or sequencing error, then it was excluded from the alignment for the given species. We treated the data from previously published studies in the same fashion as data generated in our lab by filtering the raw reads through quality trimming, then assembling de novo transcriptomes using the same programs and parameters (Table 1).

Amino acid sequences were aligned using MUSCLE v3.8.31 [85]. Alignments were also subjected to manual alignment in Aliview v1.26 [86]; extraneous data were trimmed, leaving only the aligned genes. Ambiguously aligned regions were eliminated from each alignment leaving only conserved and reliably aligned regions for phylogenetic analysis using the following parameters in Gblocks v0.91b [87]: -t=p -b3=8 -b4=2 -b5=h -b6=y. Phyutility v2.7.1 [88] was used to concatenate all gene alignment files.

Single-gene alignments were subjected to ML and BI analyses in order to infer single-gene phylogenies. Single-gene phylogenies were then further analyzed using a coalescent-based approach. The concatenated multi-gene alignment was partitioned so that the appropriate model of protein substitution was applied to each gene for the supermatrix phylogenetic approach under ML and BI.

The ML and BI analyses of the concatenated dataset used a partitioning strategy where the best evolutionary model for each gene was predicted by ProtTest v3.4.2 under the Akaike Information Criterion (AIC). For information regarding each predicted evolutionary model, please refer to Additional file 1: Table S4 [84, 89–91]. The ML analysis was conducted using IQtree v1.6.12 [92] under partition models [93]. Support values reported for the IQtree ML analysis were estimated through the bootstrap technique where 1000 ultrafast bootstrap replicates were generated [94]. The BI analysis was performed with MrBayes 3.2.7a [95] with 3 heated and 1 cold Markov chains, where trees were sampled every 1000 generations for a total of 1,000,000 generations with 1000 trees discarded at the beginning of each chain (ngen = 100000000, samplefreq = 1000, burnin = 1000, nruns = 4, nchains = 4, starttree = random).

ASTRAL [96] was used to perform the coalescent-based analysis where all 40 single-gene phylogenies produced by IQtree were used as the input after collapsing branches with low bootstrap support (< 10) using Newick Utilities v1.6 [97]. Posterior probabilities were assessed for the Bayesian and coalescent-based analyses in MrBayes and ASTRAL, respectively. Lastly, approximately unbiased (AU) tests with 100,000 RELL resamplings were conducted to test certain key topologies and hypotheses using IQtree (-zw 100000 -au) (Additional file 1: Fig. S1).

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12915-021-01087-0>.

Additional file 1 Table S1. Accession numbers of previously published RNA-seq data. **Table S2.** Information on sampled genera regarding cellularity, typical cell number, differentiation, and gamete size. **Table S3.**

Medium used to culture each sequenced strain. **Table S4.** 40 genes with best predicted evolutionary model under the Akaike information criterion (AIC). **Table S5.** National Center for Biotechnology Institute (NCBI) accession numbers for BioProject PRJNA701495. **Fig. S1.** Approximately Unbiased (AU) tests comparing key hypotheses for our 40-protein concatenated dataset. **Fig. S2.** (A) Phylogeny that represents a replication of the Zhang et al. [51] results as they relate to the volvocine algae. (B) Phylogeny of the volvocine algae that represents a change in the branching order once more volvocine taxa are sampled for phylogenetic inference. **Fig. S3.** Phylogeny of the volvocine algae which highlights the four sections of genus *Volvox* as recognized by Nozaki et al. [35].

Additional file 2 Additional methods. Provides additional detail for methods used during RNA extractions, and of how we confirmed that *Tetraabaena socialis* N-571 and N-691 are conspecific.

Additional file 3. Python script. Python script that was used to mine individual de novo transcriptomes for single-copy genes located through local BLAST databases. This script reads BLAST input files in XML format and extracts and translates a scaffold in all six reading frames based on the scaffold ID of the first BLAST hit. Once the scaffold is translated in all six reading frames, the 'hit sequence' is located and written to a file with the 'query sequence' in FASTA format.

Acknowledgements

The authors gratefully acknowledge the core facilities at the Parker H. Petit Institute for Bioengineering and Bioscience at the Georgia Institute of Technology for the use of their shared equipment, services, and expertise. We are also grateful for useful discussion with and comments on the manuscript provided by Andy Chea, Emily Cook, Sa Geng, Armin Hallmann, Joseph Lachance, Bradley Olson, and James Umen. We thank Olivier De Clerck for kindly providing amino acid alignments. This material is based upon work while Herron was serving at the National Science Foundation.

Authors' contributions

Performed the experiments (CRL), carried out the analyses (CRL, MDH), and wrote the manuscript (CRL, MDH, RFR). The authors have read and approved the final manuscript.

Funding

This work used the Hive cluster, which is supported by the National Science Foundation under grant number OAC-1828187. This research was supported in part through research cyberinfrastructure resources and services provided by the Partnership for an Advanced Computing Environment (PACE) at the Georgia Institute of Technology, Atlanta, Georgia, USA. This project was funded by NASA Astrobiology Grant# NNA17BB05A to Rosenzweig (PI) Herron (co-PI); NSF DEB-1723293 to Herron (PI) Rosenzweig (co-PI); Rosenzweig NASA Exobiology Grant # 80NSSC20K0621 to Rosenzweig (PI); Rosenzweig Georgia Tech start-up account DE00000308.

Availability of data and materials

All raw data generated and used for this study have been deposited in the National Center for Biotechnology Information (NCBI) Sequence Read Archive (SRA) under BioProject PRJNA701495 [98]. Accession numbers for our raw RNA-Seq reads range from SRR13719233 to SRR13719287, and accession numbers for our assembled contigs have been provided in Additional file 1: Table S4. For detailed information regarding accession number assignment to a specific taxon, please refer to Additional file 1: Table S5. Files containing our single-gene phylogenies and amino acid alignments, our IQtree partition file, and MrBayes configuration file have been uploaded to Dryad [99]. Previously published data used in this study from Hu et al. [47] and Featherston et al. [21] can be found under BioProject number PRJNA532307 [100] and PRJNA393411 [101], respectively.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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Received: 13 April 2021 Accepted: 8 July 2021

Published online: 31 August 2021

References

- Grosberg RK, Strathmann RR. The evolution of multicellularity: a minor major transition? *Annu Rev Ecol Syst.* 2007;38(1):621–54. <https://doi.org/10.1146/annurev.ecolsys.36.102403.114735>.
- Maynard Smith J, Szathmáry E. The major transitions in evolution: Oxford University Press; 1995.
- Szathmáry E, Maynard Smith J. The major evolutionary transitions. *Nature.* 1995;374(6519):227–32. <https://doi.org/10.1038/374227a0>.
- West SA, Fisher RM, Gardner A, Kiers ET. Major evolutionary transitions in individuality. *Proc Natl Acad Sci.* 2015;112(33):10112–9. <https://doi.org/10.1073/pnas.1421402112>.
- Hallmann A. Evolution of reproductive development in the volvocine algae. *Sex Plant Reprod.* 2011;24(2):97–112. <https://doi.org/10.1007/s00497-010-0158-4>.
- Wahl ME, Murray AW. Multicellularity makes somatic differentiation evolutionarily stable. *Proc Natl Acad Sci.* 2016;113(30):8362–7. <https://doi.org/10.1073/pnas.1608278113>.
- Chen L, Xiao S, Pang K, Zhou C, Yuan X. Cell differentiation and germ–soma separation in Ediacaran animal embryo-like fossils. *Nature.* 2014;516(7530):238–41. <https://doi.org/10.1038/nature13766>.
- Han T, Runnegar B. Megascopic eukaryotic algae from the 2.1-billion-year-old neogauee iron-formation, Michigan. *Science.* 1992;257(5067):232–5. <https://doi.org/10.1126/science.1631544>.
- Schirmer BE, de Vos JM, Antonelli A, Bagheri HC. Evolution of multicellularity coincided with increased diversification of cyanobacteria and the Great Oxidation Event. *Proc Natl Acad Sci.* 2013;110(5):1791–6. <https://doi.org/10.1073/pnas.1209927110>.
- Herron MD, Borin JM, Boswell JC, Walker J, Chen I-CK, Knox CA, et al. De novo origins of multicellularity in response to predation. *Sci Rep.* 2019;9(1):2328. <https://doi.org/10.1038/s41598-019-39558-8>.
- Quintero-Galvis JF, Paleo-López R, Solano-Igauran JJ, Poupin MJ, Ledger T, Gaitan-Espitia JD, et al. Exploring the evolution of multicellularity in *Saccharomyces cerevisiae* under bacteria environment: an experimental phylogenetics approach. *Ecol Evol.* 2018;8(9):4619–30. <https://doi.org/10.1002/ece3.3979>.
- Ratcliff WC, Denison RF, Borrello M, Travisano M. Experimental evolution of multicellularity. *Proc Natl Acad Sci.* 2012;109(5):1595–600. <https://doi.org/10.1073/pnas.1115323109>.
- Ratcliff WC, Herron MD, Howell K, Pentz JT, Rosenzweig F, Travisano M. Experimental evolution of an alternating uni- and multicellular life cycle in *Chlamydomonas reinhardtii*. *Nat Commun.* 2013;4:1–7.
- Herron MD, Michod RE. Evolution of complexity in the volvocine algae: transitions in individuality through Darwin's eye. *Evolution.* 2008;62(2):436–51. <https://doi.org/10.1111/j.1558-5646.2007.00304.x>.
- Kirk DL. A twelve-step program for evolving multicellularity and a division of labor. *BioEssays.* 2005;27(3):299–310. <https://doi.org/10.1002/bies.20197>.
- Koyanagi KO. Inferring cell differentiation processes based on phylogenetic analysis of genome-wide epigenetic information: hematopoiesis as a model case. *Genome Biol Evol.* 2015;7(3):699–705. <https://doi.org/10.1093/gbe/evv024>.
- van Leeuwenhoek A. Part of a letter from Mr Antony van Leeuwenhoek, concerning the worms in sheeps livers, gnats, and animalcula in the excrements of frogs. *Philos Trans R Soc Lond.* 1700;22:509–18.
- Kirk DL. Germ–soma differentiation in *Volvox*. *Dev Biol.* 2001;238(2):213–23. <https://doi.org/10.1006/dbio.2001.0402>.
- Schmitt R. Differentiation of germinal and somatic cells in *Volvox carteri*. *Curr Opin Microbiol.* 2003;6(6):608–13. <https://doi.org/10.1016/j.mib.2003.10.007>.
- Craig RJ, Hasan AR, Ness RW, Keightley PD. Comparative genomics of *Chlamydomonas*. *Plant Cell.* 2021;33(4):1016–41. <https://doi.org/10.1093/plcell/koab026>.
- Featherston J, Arakaki Y, Hanschen ER, Ferris PJ, Michod RE, Olson BJSC, et al. The 4-celled *Tetrabaena socialis* nuclear genome reveals the essential components for genetic control of cell number at the origin of multicellularity in the volvocine lineage. *Mol Biol Evol.* 2018;35(4):855–70. <https://doi.org/10.1093/molbev/msx332>.
- Hanschen ER, Marriage TN, Ferris PJ, et al. The *Gonium pectorale* genome demonstrates co-option of cell cycle regulation during the evolution of multicellularity. *Nat Commun.* 2016;7:1–10.
- Merchant SS, Prochnik SE, Vallon O, Harris EH, Karpowicz SJ, Witman GB, et al. The *Chlamydomonas* genome reveals the evolution of key animal and plant functions. *Science.* 2007;318(5848):245–50. <https://doi.org/10.1126/science.1143609>.
- Prochnik SE, Umen J, Nedelcu AM, Hallmann A, Miller SM, Nishii I, et al. Genomic analysis of organismal complexity in the multicellular green alga *Volvox carteri*. *Science.* 2010;329(5988):223–6. <https://doi.org/10.1126/science.1188800>.
- Herron MD, Hackett JD, Aylward FO, Michod RE. Triassic origin and early radiation of multicellular volvocine algae. *Proc Natl Acad Sci.* 2009;106(9):3254–8. <https://doi.org/10.1073/pnas.0811205106>.
- Brunet T, King N. The origin of animal multicellularity and cell differentiation. *Dev Cell.* 2017;43(2):124–40. <https://doi.org/10.1016/j.devcel.2017.09.016>.
- Lang NJ. Electron microscopy of the Volvocaceae and Astrephomenaceae. *Am J Bot.* 1963;50(3):280–300. <https://doi.org/10.1002/j.1537-2197.1963.tb12235.x>.
- Larson A, Kirk MM, Kirk DL. Molecular phylogeny of the volvocine flagellates. *Mol Biol Evol.* 1992;9(1):85–105. <https://doi.org/10.1093/oxfordjournals.molbev.a040710>.
- Hanschen ER, Herron MD, Wiens JJ, Nozaki H, Michod RE. Multicellularity drives the evolution of sexual traits. *Am Nat.* 2018;192(3):E93–E105. <https://doi.org/10.1086/698301>.
- Herron MD, Desnitskiy AG, Michod RE. Evolution of developmental programs in *Volvox* (Chlorophyta). *J Phycol.* 2010;46(2):316–24. <https://doi.org/10.1111/j.1529-8817.2009.00803.x>.
- Hanschen ER, Herron MD, Wiens JJ, Nozaki H, Michod RE. Repeated evolution and reversibility of self-fertilization in the volvocine green algae*. *Evolution.* 2018;72(2):386–98. <https://doi.org/10.1111/evo.13394>.
- Nakada T, Tomita M, Nozaki H. *Volvulina compacta* (Volvocaceae, Chlorophyceae), new to Japan, and its phylogenetic position. *J Jpn Bot.* 2010;85:364–9.
- Nozaki H, Misawa K, Kajita T, Kato M, Nohara S, Watanabe MM. Origin and evolution of the colonial Volvocales (Chlorophyceae) as inferred from multiple, chloroplast gene sequences. *Mol Phylogenet Evol.* 2000;17(2):256–68. <https://doi.org/10.1006/mpev.2000.0831>.
- Nozaki H, Yamada TK, Takahashi F, Matsuzaki R, Nakada T. New “missing link” genus of the colonial volvocine green algae gives insights into the evolution of oogamy. *BMC Evol Biol.* 2014;14:1–11.
- Nozaki H, Matsuzaki R, Yamamoto K, Kawachi M, Takahashi F. Delineating a new heterothallic species of *Volvox* (Volvocaceae, Chlorophyceae) using new strains of “*Volvox africanus*.”. *PLoS ONE.* 2015;10:e0142632.
- Grochau-Wright ZI, Hanschen ER, Ferris PJ, Hamaji T, Nozaki H, Olson BJSC, et al. Genetic basis for soma is present in undifferentiated volvocine green algae. *J Evol Biol.* 2017;30(6):1205–18. <https://doi.org/10.1111/jeb.13100>.
- Pröschold T, Darienko T, Krienitz L, Coleman AW. *Chlamydomonas schloesseri* sp. nov. (Chlamydomonadales, Chlorophyta) revealed by morphology, autolysin cross experiments, and multiple gene analyses. *Phytotaxa.* 2018;362(1):21–38. <https://doi.org/10.11646/phytotaxa.362.1.2>.
- Nakada T, Tsuchida Y, Tomita M. Improved taxon sampling and multigene phylogeny of unicellular chlamydomonads closely related to the colonial volvocine lineage Tetrabaenaceae-Goniaceae-Volvocaceae (Volvocales, Chlorophyceae). *Mol Phylogenet Evol.* 2019;130:1–8. <https://doi.org/10.1016/j.jmpev.2018.09.013>.
- Coleman AW. Phylogenetic analysis of “Volvocaceae” for comparative genetic studies. *Proc Natl Acad Sci.* 1999;96(24):13892–7. <https://doi.org/10.1073/pnas.96.24.13892>.
- Hu Y, Xing W, Song H, Zhu H, Liu G, Hu Z. Evolutionary analysis of unicellular species in chlamydomonadales through chloroplast genome comparison with the colonial volvocine algae. *Front Microbiol.* 2019;10:1351. <https://doi.org/10.3389/fmicb.2019.01351>.
- Dong W, Liu J, Yu J, Wang L, Zhou S. Highly variable chloroplast markers for evaluating plant phylogeny at low taxonomic levels and for DNA barcoding. *PLoS ONE.* 2012;7(4):e35071. <https://doi.org/10.1371/journal.pone.0035071>.

42. Jakob SS, Blattner FR. A chloroplast genealogy of *Hordeum* (Poaceae): long-term persisting haplotypes, incomplete lineage sorting, regional extinction, and the consequences for phylogenetic inference. *Mol Biol Evol.* 2006;23(8):1602–12. <https://doi.org/10.1093/molbev/msl018>.
43. Xu B, Wu N, Gao X-F, Zhang L-B. Analysis of DNA sequences of six chloroplast and nuclear genes suggests incongruence, introgression, and incomplete lineage sorting in the evolution of *Lespedeza* (Fabaceae). *Mol Phylogenet Evol.* 2012;62(1):346–58. <https://doi.org/10.1016/j.ympev.2011.10.007>.
44. Rose J, Toledo CA, Lemmon EM, Lemmon AR, Systsma K. Out of sight, out of mind: widespread nuclear and plastid-nuclear discordance in the flowering plant genus *Polemonium* (Polemoniaceae) suggests widespread historical gene flow despite limited nuclear signal. *Syst Biol.* 2020;70:162–80.
45. Soltis DE, Kuzoff RK. Discordance between nuclear and chloroplast phylogenies in the *Heuchera* group (Saxifragaceae). *Evolution.* 1995;49(4):727–42. <https://doi.org/10.1111/j.1558-5646.1995.tb02309.x>.
46. Yu W-B, Huang P-H, Li D-Z, Wang H. Incongruence between nuclear and chloroplast DNA phylogenies in *Pedicularis* section *Cyathophora* (Orobanchaceae). *PLoS ONE.* 2013;8(9):e74828. <https://doi.org/10.1371/journal.pone.0074828>.
47. Hu Y, Xing W, Song H, Hu Z, Liu G. Comparison of colonial volvocine algae based on phylotranscriptomic analysis of gene family evolution and natural selection. *Eur J Phycol.* 2020;55(1):100–12. <https://doi.org/10.1080/09670262.2019.1663269>.
48. Nakazawa A, Krienitz L, Nozaki H. Taxonomy of the unicellular green algal genus *Vitreochlamys* (Volvocales), based on comparative morphology of cultured material. *Eur J Phycol.* 2001;36(2):113–28. <https://doi.org/10.1080/09670260110001735268>.
49. Nozaki H. Morphology and evolution of sexual reproduction in the Volvocaceae (Chlorophyta). *J Plant Res.* 1996;109(3):353–61. <https://doi.org/10.1007/BF02344484>.
50. Nozaki H, Itoh M. Phylogenetic relationships within the colonial Volvocales (Chlorophyta) inferred from cladistic analysis based on morphological data. *J Phycol.* 1994;30(2):353–65. <https://doi.org/10.1111/j.0022-3646.1994.00353.x>.
51. Zhang Z, Qu C, Yao R, Nie Y, Xu C, Miao J, et al. The parallel molecular adaptations to the Antarctic cold environment in two psychrophilic green algae. *Genome Biol Evol.* 2019;11(7):1897–908. <https://doi.org/10.1093/gbe/evz104>.
52. Nozaki H, Ohtani S. *Gonium sociale* (Volvocales, Chlorophyta) from Antarctica. *Jpn J Phycol.* 1992;40:267–71.
53. Mai JC, Coleman AW. The internal transcribed spacer 2 exhibits a common secondary structure in green algae and flowering plants. *J Mol Evol.* 1997;44(3):258–71. <https://doi.org/10.1007/PL00006143>.
54. Hedtke SM, Townsend TM, Hillis DM. Resolution of phylogenetic conflict in large data sets by increased taxon sampling. *Syst Biol.* 2006;55(3):522–9. <https://doi.org/10.1080/10635150600697358>.
55. Pollock DD, Zwickl DJ, McGuire JA, Hillis DM. Increased taxon sampling is advantageous for phylogenetic inference. *Syst Biol.* 2002;51(4):664–71. <https://doi.org/10.1080/10635150290102357>.
56. Zwickl DJ, Hillis DM. Increased taxon sampling greatly reduces phylogenetic error. *Syst Biol.* 2002;51(4):588–98. <https://doi.org/10.1080/10635150290102339>.
57. Nozaki H, Itoh M, Watanabe MM, Kuroiwa T. Ultrastructure of the vegetative colonies and systematic position of *Basichlamys* (Volvocales, Chlorophyta). *Eur J Phycol.* 1996;31(1):67–72. <https://doi.org/10.1080/09670269600651211a>.
58. Nozaki H, Ito M, Uchida H, Watanabe MM, Takahashi H, Kuroiwa T. Phylogenetic analysis of *Yamagishiella* and *Platydorina* (volvocaceae, chlorophyta) based on rbcL gene sequences. *J Phycol.* 1997;33(2):272–8. <https://doi.org/10.1111/j.0022-3646.1997.00272.x>.
59. Nozaki H, Itoh M, Sano R, Uchida H, Watanabe MM, Kuroiwa T. Phylogenetic relationships within the colonial volvocales (chlorophyta) inferred from rbcL gene sequence data. *J Phycol.* 1995;31(6):970–9. <https://doi.org/10.1111/j.0022-3646.1995.00970.x>.
60. Nozaki H. Ultrastructure of the extracellular matrix of *Gonium* (Volvocales, Chlorophyta). *Phycologia.* 1990;29(1):1–8. <https://doi.org/10.2216/i0031-8884-29-1-1.1>.
61. Nozaki H, Kuroiwa T. Ultrastructure of the extracellular matrix and taxonomy of *Eudorina*, *Pleodorina* and *Yamagishiella* gen. nov. (Volvocaceae, Chlorophyta). *Phycologia.* 1992;31(6):529–41. <https://doi.org/10.2216/i0031-8884-31-6-529.1>.
62. Kirk DL, Birchem R, King N. The extracellular matrix of *Volvox*: a comparative study and proposed system of nomenclature. *J Cell Sci.* 1986;80(1):207–31. <https://doi.org/10.1242/jcs.80.1.207>.
63. Herron MD. Origins of multicellular complexity: *Volvox* and the volvocine algae. *Mol Ecol.* 2016;25(6):1213–23. <https://doi.org/10.1111/mec.13551>.
64. da Silva J, Drysdale VL. Isogamy in large and complex volvocine algae is consistent with the gamete competition theory of the evolution of anisogamy. *Proc R Soc B Biol Sci.* 2018;285(1890):20181954. <https://doi.org/10.1098/rspb.2018.1954>.
65. Umen JG. Evolution of sex and mating loci: an expanded view from Volvocine algae. *Curr Opin Microbiol.* 2011;14(6):634–41. <https://doi.org/10.1016/j.mib.2011.10.005>.
66. Coleman AW. Biogeography and speciation in the *Pandorina/Volvulina* (chlorophyta) superclade. *J Phycol.* 2001;37(5):836–51. <https://doi.org/10.1046/j.1529-8817.2001.01043.x>.
67. Nozaki H. Origin and evolution of the genera *Pleodorina* and *Volvox* (Volvocales). *Biologia.* 2003;58:425–31.
68. Nozaki H, Ott FD, Coleman AW. Morphology, molecular phylogeny and taxonomy of two new species of *Pleodorina* (Volvocales, Chlorophyceae). *J Phycol.* 2006;42(5):1072–80. <https://doi.org/10.1111/j.1529-8817.2006.00255.x>.
69. Smith GM. A comparative study of the species of *Volvox*. *Trans Am Microsc Soc.* 1944;63(4):265–310. <https://doi.org/10.2307/3223302>.
70. Nozaki H, Ohta N, Takano H, Watanabe MM. Reexamination of phylogenetic relationships within the colonial Volvocales (Chlorophyta): an analysis of *atpB* and *rbcL* gene sequences. *J Phycol.* 1999;35(1):104–12. <https://doi.org/10.1046/j.1529-8817.1999.3510104.x>.
71. Nakada T, Misawa K, Nozaki H. Molecular systematics of Volvocales (Chlorophyceae, Chlorophyta) based on exhaustive 18S rRNA phylogenetic analyses. *Mol Phylogenet Evol.* 2008;48(1):281–91. <https://doi.org/10.1016/j.ympev.2008.03.016>.
72. Pröschold T, Marin B, Schlösser UG, Melkonian M. Molecular phylogeny and taxonomic revision of *Chlamydomonas* (Chlorophyta). I. Emendation of *Chlamydomonas* Ehrenberg and *Chloromonas* Gobi, and description of *Oogamochlamys* gen. nov. and *Lobochlamys* gen. nov. *Protist.* 2001;152:36.
73. Kato S. Laboratory culture and morphology of *Colacium vesiculosum* Ehrb. (Euglenophyceae). *Jpn J Phycol.* 1982;30:63–7.
74. Ichimura T, Watanabe M. The *Closterium calosporum* complex from the Ryukyu Islands - variation and taxonomical problems. *Mem Natn Sci Mus Tokyo.* 1974. p. 7.
75. Ichimura T (1973) The life cycle and its control in some species of *Closterium*, with special reference to the biological species problems. *Dr. Diss.*
76. Starr EC. Special methods-dry soil samples. In: Stein JR, editor. Cambridge: Cambridge University Press; 1973.
77. Sueoka N. Mitotic replication of deoxyribonucleic acid in *Chlamydomonas reinhardtii*. *Natl Acad Sci U S Am.* 1960;46(1):83–91. <https://doi.org/10.1073/pnas.46.1.83>.
78. Matt GY, Umen JG. Cell-type transcriptomes of the multicellular green alga *Volvox carterii* yield insights into the evolutionary origins of germ and somatic differentiation programs. *G3amp58 GenesGenomesGenetics.* 2018;8:531–50.
79. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics.* 2014;30(15):2114–20. <https://doi.org/10.1093/bioinformatics/btu170>.
80. Xie Y, Wu G, Tang J, Luo R, Patterson J, Liu S, et al. SOAPdenovo-Trans: de novo transcriptome assembly with short RNA-Seq reads. *Bioinformatics.* 2014;30(12):1660–6. <https://doi.org/10.1093/bioinformatics/btu077>.
81. Fu L, Niu B, Zhu Z, Wu S, Li W. CD-HIT: accelerated for clustering the next-generation sequencing data. *Bioinformatics.* 2012;28(23):3150–2. <https://doi.org/10.1093/bioinformatics/bts565>.
82. Hasegawa M, Hashimoto T. Ribosomal RNA trees misleading? *Nature.* 1993;361(6407):23. <https://doi.org/10.1038/361023b0>.
83. Loomis WF, Smith DW. Molecular phylogeny of *Dictyostelium discoideum* by protein sequence comparison. *Proc Natl Acad Sci.* 1990;87(23):9093–7. <https://doi.org/10.1073/pnas.87.23.9093>.
84. De Clerck O, Kao S-M, Bogaert KA, et al. Insights into the evolution of multicellularity from the sea lettuce genome. *Curr Biol.* 2018;28:2921–2933.e5.
85. Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 2004;32(5):1792–7. <https://doi.org/10.1093/nar/gkh340>.
86. Larsson A. AliView: a fast and lightweight alignment viewer and editor for large datasets. *Bioinformatics.* 2014;30(22):3276–8. <https://doi.org/10.1093/bioinformatics/btu531>.
87. Castresana J. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Mol Biol Evol.* 2000;17(4):540–52. <https://doi.org/10.1093/oxfordjournals.molbev.a026334>.

88. Smith SA, Dunn CW. Phyutility: a phyloinformatics tool for trees, alignments and molecular data. *Bioinformatics*. 2008;24(5):715–6. <https://doi.org/10.1093/bioinformatics/btm619>.
89. Le SQ, Gascuel O. An improved general amino acid replacement matrix. *Mol Biol Evol*. 2008;25(7):1307–20. <https://doi.org/10.1093/molbev/msn067>.
90. Jones DT, Taylor WR, Thornton JM. The rapid generation of mutation data matrices from protein sequences. *Bioinformatics*. 1992;8(3):275–82. <https://doi.org/10.1093/bioinformatics/8.3.275>.
91. Dayhoff MO, Schwartz RM, Orcutt BC. A model of evolutionary change in proteins. *Atlas Protein Seq Struct*. 1978;5:345–52.
92. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol*. 2015;32(1):268–74. <https://doi.org/10.1093/molbev/msu300>.
93. Chernomor O, von Haeseler A, Minh BQ. Terrace aware data structure for phylogenomic inference from supermatrices. *Syst Biol*. 2016;65(6):997–1008. <https://doi.org/10.1093/sysbio/syw037>.
94. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. UFBoot2: improving the ultrafast bootstrap approximation. *Mol Biol Evol*. 2018;35(2):518–22. <https://doi.org/10.1093/molbev/msx281>.
95. Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, et al. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol*. 2012;61(3):539–42. <https://doi.org/10.1093/sysbio/sys029>.
96. Zhang C, Rabiee M, Sayyari E, Mirarab S. ASTRAL-III: polynomial time species tree reconstruction from partially resolved gene trees. *BMC Bioinform*. 2018;19(S6):153. <https://doi.org/10.1186/s12859-018-2129-y>.
97. Junier T, Zdobnov EM. The Newick utilities: high-throughput phylogenetic tree processing in the UNIX shell. *Bioinformatics*. 2010;26(13):1669–70. <https://doi.org/10.1093/bioinformatics/btq243>.
98. Lindsey C, Rosenzweig F, Herron MD. Phylotranscriptomics points to multiple independent origins of multicellularity and cellular differentiation in the volvocine algae. *NCBI BioProjects*. <https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA701495>.
99. Lindsey C, Rosenzweig F, Herron MD. Phylotranscriptomics points to multiple independent origins of multicellularity and cellular differentiation in the volvocine algae. *Dryad*. <https://doi.org/10.5061/dryad.v9s4mw6w5>.
100. Hu Y, Xing W, Song H, Hu Z, Liu G. Comparison of colonial volvocine algae based on phylotranscriptomic analysis of gene family evolution and natural selection. *NCBI BioProjects*. <https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA532307>.
101. Featherston J, Arakaki Y, Hanschen ER, Ferris PJ, Michod RE, Olson BJSC, et al. The 4-celled *Tetraabaena socialis* nuclear genome reveals the essential components for genetic control of cell number at the origin of multicellularity in the volvocine lineage. *NCBI BioProjects*. <https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA393411>.

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